

Models for Insulin Dependent Diabetes Mellitus in Humans

Abstract

Insulin-dependent diabetes mellitus is generally considered to result from a T-cell mediated autoimmune destruction of the insulin producing β -cells in the pancreatic islets of Langerhans. Considerable progress has been made with regard to the unraveling of the pathogenesis of the disease, but neither the initiation of the disease process, nor the progression to overt insulin-dependency is understood. A major advantage in case of type 1 diabetes is the presence of spontaneous animal models of the disease that allow investigation of particular aspects of the disease. While in most autoimmune diseases, the process must be initiated, either by breakdown of tolerance through chemicals or induction of autoimmunity with specific autoantigens, diabetes develops spontaneously in non-obese diabetic (NOD) mice and biobreeding (BB) rats. Despite this major advantage, both models have considerable limitations. The present article deals with this notion, with focus on the best studied model in NOD mice. It is concluded that, although much can be learned from animal models, extrapolation to human disease must be done with great care. To put it extreme, an inbred mouse or rat strain can be considered as a case-report of the disease in humans.

Genetics

Type 1 diabetes in humans is characterized by a relatively strong but heterogeneous genetic predisposition, with a polygenic trait (1-3). In particular genetic loci within the major histocompatibility complex on the short arm of chromosome 6 are strongly associated with either susceptibility to, or protection against development of diabetes. Interestingly, the major contributor, designated IDDM-1, consists of several HLA loci within the MHC region (4, 5). In Table 1, the frequencies of certain HLA class II alleles are indicated in a large cohort of juvenile onset type 1 diabetes in the Netherlands. From these data, it is evident that, although certain combinations of haplotypes of HLA-DR and -DQ are clearly far more frequently present amongst IDDM patients than in the general population, less than half of these patients have the predisposing haplotypes, while 11% of the general population contains these high-risk haplotypes. This implies that the vast majority of subjects with high risk

Table 1. Comparison of three most frequent HLA phenotypes in IDDM patients with general population.

HLA	IDDM patients (n=309)	General population (dutch) (N=504)
DR3/3 – DQ2/2	8.1%	3.0%
DR3/4 – DQ2/8	30.7%	3.4%
DR4/4 – DQ8/8	5.8%	0.2%
<i>High risk total</i>	<i>44.6%</i>	<i>6.6%</i>
<i>Low risk (DR2 – DQ6)</i>	<i>0.9%</i>	<i>25.4%</i>

haplotypes will never develop IDDM, since the incidence rates in the population at large are in the range of 0.1-0.4%. In families with IDDM, first degree relatives have an increased risk (ca. 5%), while identical twins show concordance rates of 25-55%. Thus, HLA genes are important, but neither required nor sufficient for the development of IDDM. Moreover, diabetes specific genes or alleles have not been identified. This suggests that particular combinations of predisposing genetic loci, rather than individual genes, will determine the actual risk of disease development. The results of the genetic studies also imply that environmental factors strongly influence the genetic trait.

While animal models are useful to study the genetic predisposition to diabetes, environmental issues associated with disease development in humans cannot easily be addressed in animals.

Genetic predisposition is also associated with disease progression in NOD mice (6-8). Again, a polygenic trait is evident. Multiple genetic loci contribute to development of diabetes, but as in humans, MHC is most strongly associated with the disease (Table 2). The similarities with diabetes in humans is even more striking with regard to protection. Aspartic acid on position 57 of the DQB chain (e.g. DQB1*0602) in men (9), or mice (10) is associated with strong protection against development of diabetes. However, while a wide range of HLA class II alleles are associated with susceptibility to diabetes in humans, the mouse equivalent I-A^{g7} is the only MHC haplotype in mice to be associated with this disease. Nonetheless, several non-MHC regions contributing to disease predisposition appear to be homologous between mice and men.

Immunology

Type 1 diabetes in humans is characterized by rather general immunological phenotypes. A defective peripheral immunoregulation has been described in several studies (11-15). Even at the level of CD4/CD8 ratios, differences between patients and controls are reported. Few studies on prediabetic subjects have been published, but the data suggest that abnormalities on the levels of lymphocyte subsets exist prior to clinical presentation of diabetes. The relative increases in T-cells expressing both CD45RA and CD45RO, which represent in vivo activated T-cells as illustrated by the expression of HLA class II and CD25, may be caused by a maturation defect in

Table 2. Comparison of insulin-dependent diabetes in men and mice.

	Humans	NOD mice
Genetic predisposition, polygenic trait	Yes	Yes
IDDM1/ <i>idd-1</i>	Multiple haplotypes	One haplotype
Environmental effects	Probable	Yes
Endogenous retrovirus in β -cells	?	Yes
Incidence	0.3%	80-90%
Age at diabetes onset	0 – >70 years	3 months
Gender bias	No	Yes
Defective peripheral regulation	Yes	Yes
T-cell driven insulinitis	Mild	Severe
Peri-insulinitis	(no)	Yes
Leukocyte infiltrates in other tissues	sometimes	Always
Disease transmission via BMT	Yes	Yes
Humoral reactivity to β -cells	yes	(yes)
Autoantigens	GAD65, insulin, IA-2, 38kD, ...	GAD65, insulin, 38kD, peripherin, ...
Delayed onset with immunosuppression	Yes	Yes
Successful intervention therapies	?	OKT3, anti-MHC, intra-oral, -venous or -nasal autoantigens and peptides

diabetic patients (16). Alternatively, a constant priming, e.g. with islet autoantigens or viral products, may also result in the relative increase of activated lymphocytes (15). It is generally believed that the cytokine production pattern of pathogenic T-cells is Th1-like (i.e. interferon- γ , IL-2, TNF), while non-destructive T-cell autoreactivity appears to be anti-inflammatory in nature (Th2; IL-4, IL-5 and IL-10) (17). This dichotomy is primarily based on animal studies (18, 19), but most studies on cytokine production of T-cells reactive with islet cell autoantigen support this concept (20, 21).

All evidence so far supports a T-cell driven pathogenesis of type 1 diabetes in humans. In addition to the abnormalities described above, studies using immunosuppressive agents (22, 23), as well as case reports on adoptive transfer with non-T-cell-depleted bone marrow from a diabetic patient to an immuno-incompetent relative of this patient (24) are in line with this hypothesis. In fact, recurrent insulinitis and β -cell destruction that was noted in patients receiving a pancreas segment of their non-diabetic twin was observed in the absence of islet autoantibodies (25). The latter are commonly used to predict onset of IDDM (26). Moreover, islet cell autoantibodies have been used to define the potential targets of pathogenic T-cells in IDDM. Thus far, GAD65, insulin, IA-2 and ICA69 have been defined on this basis (27-32), while other candidates were defined by T-cells, rather than autoantibodies (imogen-38, insulin-secretory granule protein of 38kDa) (33, 34). Thus far, none of these candidates were defined primarily on basis of animal models of diabetes. In fact, presence of various islet autoantibodies in NOD mice is still disputed.

The NOD mouse model has contributed significantly to the design of studies to define the pathogenesis of diabetes in humans. The notion that T-cells play a central role in β -cell destruction were corroborated with the finding of insulitis in men (35) and mice (36), although the so-called non-destructive insulitis or peri-insulitis (36-40) has not been reported in humans so far (Table 2). Furthermore, the degree and extent of insulitis in humans is much less than that in NOD mice (41, 42). Interestingly, all NOD mice will develop a degree of insulitis, regardless of lack of progression to overt diabetes.

Other studies to support the role of T-cells in NOD mice include adoptive transfer (43-45), lack of diabetes in athymic mice and prevention of disease with monoclonal antibodies against T-cells (46). While several candidate autoantigens are recognized by T-cells of mice and men, such as GAD65 (28, 47), insulin (30, 48, 49) and insulin-secretory granule protein 38kDa (34, 50, 51), others have only been reported in either mice (peripherin (52), hsp60 (53)) or men (IA-2 (54)).

Recently, HLA-DR and -DQ transgenic mice have been used to study HLA restricted T-cell reactivity to human islet autoantigens *in vivo*. Interestingly, the immunodominant HLA-restricted epitopes that were defined after immunization with human autoantigens were identical to those identified by human T-cell clones isolated from type 1 diabetes patients (55-60). In fact, the HLA binding affinity of T-cell epitopes defined upon immunization of HLA transgenic mice and patient derived T-cells was relatively high, while epitopes of T-cells generated from non-diabetic subjects was relatively low (59). These findings implicate that in this context, mouse studies may be useful to study *in vivo* reactivity of HLA restricted T-cells specific to human autoantigen and the definition of human T-cell epitopes.

Immunotherapy

The experience with immunotherapy is relatively limited in human type 1 diabetes. Several trials using immunosuppressive agents have been applied with varying degrees of clinical benefit, but it is evident that general immunosuppression potentially delays the clinical onset of diabetes, or increases the rate of remission. The list of immunointervention trials in diabetes is long (61-63). At this stage, most emphasis is put on nicotinamide (64), insulin prophylaxis (37, 65) and oral or nasal tolerance induction with insulin (66-68). Clearly, studies in mice are a logical first step to assess efficacy of treatment, although failure in mice does not preclude potential success in humans. It has been convincingly shown that T-cells to either GAD65 (47, 69), insulin (48), hsp60 (53) or 38kDa secretory granule protein (50) can cause β -cell destruction and diabetes in NOD mice. In fact, T-cells reactive to other yet undefined islet autoantigens have also been shown to be pathogenic in NOD mice (40, 70). The known candidate autoantigens have effectively been used to prevent or delay autoimmune destruction of β -cells in NOD mice (37, 68, 71-75). However, it remains to be demonstrated that these antigens indeed are suitable and effective in humans, without aggravating the autoimmune response to either β -cells (76), or other tissue, since none of the candidate autoantigens are islet specific.

Caveats

As illustrated above, the similarities between men and NOD mice are astonishing in many respects, but evident discrepancies in the pathogenic processes between these two species must be appreciated. First, the homogeneous nature of inbred mouse strains such as NOD mice is an advantage with regard to synchronization of diabetes onset, high incidence rates and limited inter-individual biological variability, while at the same time it does not pay tribute to the large heterogeneity of type 1 diabetes in humans in terms of genetic background, clinical presentation and phenotype.

Second, particular aspects of the immune process in IDDM can be studied in great detail with help of transgenic (40, 77-85) and knock-out (86) mouse models. Unfortunately, such models have obvious disadvantages once extrapolated to the situation in humans for the obvious reasons, and can introduce cell biological artifacts that obscure 'natural' tolerance and autoimmunity. On the same line, the mechanisms involved in islet graft survival or destruction may be quite different from the 'natural' pathogenesis of type 1 diabetes, both in mice and men (87-89). For instance, intrathymic implantation of islets of Langerhans have only been successful in achieving tolerance in mice and rats (90), while such efforts failed in non-rodent mammals. Finally, the interpretations of studies on adoptive transfer in NOD mice must be done with great care. Clearly, the kinetics of the β -cell destruction process are quite different, but more importantly, the treatment of the recipient mice with either irradiation or chemicals interferes with regulatory processes that occur *in vivo*. It may be argued that in fact, the mechanism of adoptive transfer is more similar to graft-versus-host reactions than spontaneous autoimmune reactivity in terms of effector cell numbers, treatment of recipients and kinetics. Nonetheless, diabetologists should be grateful to have models as close to the disease in humans as the spontaneous model in NOD mice. Surely, the mice have been very informative and helpful in shaping the experimental approaches to unravel human autoimmune β -cell destruction.

References

1. Todd, J.A., 1990. Genetic control of autoimmunity in type 1 diabetes. *Immunol. Today* **11**:122-129.
2. Svejgaard, A, B.K. Jakobsen, P. Platz, L.P. Ryder, J. Nerup and M. Christy et al., 1986. HLA associations in insulin-dependent diabetes: search for heterogeneity in different groups of patients from a homogeneous population. *Tissue Antigens* **28**:237-244.
3. Undlien, D.E., T. Friede, H.G. Rammensee, G. Joner, K. Dahl-Jorgensen and H.E. Akselsen et al., 1997. HLA-encoded genetic predisposition in IDDM: DR4 subtypes may be associated with different degrees of protection. *Diabetes* **46**:143-149.
4. Davies, J.L., Y. Kawaguchi, S.T. Bennett, J.B. Copeman, H.J. Cordell and L.E. Pritchard et al., 1994. A genome-wide search for human type 1 diabetes susceptibility genes. *Nature* **371**:130-136.
5. Hanifi Moghaddam, P., P. de Knijff, B.O. Roep, B. van der Auwera, A. Naipal, F.K. Gorus, F. Schuit, M.J. Giphart and Belgian Diabetes Registry. 1998. Genetic structure of IDDM1: Two separate regions in the Major Histocompatibility Complex contribute to susceptibility or protection. *Diabetes* **47**, 263-269.
6. Prochazka, M., D.V. Serreze, S.M. Worthen and E.H. Leiter, 1989. Genetic control of diabetogenesis in NOD/Lt mice. Development and analysis of congenic stocks. *Diabetes* **38**:1446-1455.
7. Serreze, D.V. and E.H. Leiter, 1994. Genetic and pathogenic basis of autoimmune diabetes in NOD mice. *Curr.Opin.Immunol.* **6**:900-906.

8. Ghosh, S., S.M. Palmer, N.R. Rodrigues, H.J. Cordell, C.M. Hearne, R.J. Cornall et al., 1993. Polygenic Control of Autoimmune Diabetes in Nonobese Diabetic Mice. *Nat. Genet.* **4**:404-409.
9. Todd, J.A., J.I. Bell and H.O. McDavitt, 1987. HLA-DQ beta gene contributes to susceptibility and resistance to insulin-dependent diabetes mellitus. *Nature* **329**:599-604.
10. Acha-Orbea, H. and H.O. McDavitt, 1987. The first external domain of the nonobese diabetic mouse class II I-A beta chain is unique. *Proc. Natl. Acad. Sci. USA* **84**:2435-2439.
11. Peakman, M., J.M. Tredger, E.T. Davies, M. Davenport, J.B. Dunne, R. Williams et al., 1993. Analysis of peripheral blood mononuclear cells in rodents by three-colour flow cytometry using a small-volume lysed whole blood technique. *J. Immunol. Methods* **158**:87-94.
12. Smerdon, R.A., M. Peakman, M.J. Hussain, L. Alviggi, P.J. Watkins, D.G. Leslie et al., 1993. Increase in Simultaneous Coexpression of Naive and Memory Lymphocyte Markers at Diagnosis of IDDM. *Diabetes* **42**:127-133.
13. al-Sakkaf, L., P. Pozzilli, A.C. Tarn, G. Schwarz, E.A. Gale and G.F. Bottazzo, 1989. Persistent reduction of CD4/CD8 lymphocyte ratio and cell activation before the onset of type 1 (insulin-dependent) diabetes. *Diabetologia* **32**:322-325.
14. Buschard, K., P. Damsbo and C. Ropke, 1990. Activated CD4+ and CD8+ T-lymphocytes in newly diagnosed type 1 diabetes: a prospective study. *Diabet. Med.* **7**:132-136.
15. Douglas Petersen, L., G. Duinkerken, G.J. Bruining, R. van Lier, R.R.P. De Vries and B.O. Roep, 1996. Increased numbers of in vivo activated T-cells in patients with recent onset diabetes mellitus. *J. Autoimmun.* **9**:731-737.
16. Faustman, D., G. Eisenbarth, J. Daley and J. Breitmeyer, 1989. Abnormal T-lymphocyte subsets in type I diabetes. *Diabetes* **38**:1462-1468.
17. Rabinovitch, A., 1993. Roles of cytokines in IDDM pathogenesis and islet β -cell destruction. *Diabetes Rev.* **1**:215-240.
18. Rothe, H., A. Faust, U. Schade, R. Kleemann, G. Bosse, T. Hibino et al., 1994. Cyclophosphamide treatment of female non-obese diabetic mice causes enhanced expression of inducible nitric oxide synthase and interferon-gamma, but not of interleukin-4. *Diabetologia* **37**:1154-1158.
19. Mosmann, T.R., 1992. T lymphocyte subsets, cytokines, and effector functions. [Review]. *Ann. N.Y. Acad. Sci.* **664**:89-92.
20. Harrison, L.C., M.C. Honeyman, H.J. De Aizpurua, R.S. Schmidli, P.G. Colman, B.D. Tait et al., 1993. Inverse relation between humoral and cellular immunity to glutamic acid decarboxylase in subjects at risk of insulin-dependent diabetes. *Lancet* **341**:1365-1369.
21. Kallan, A.A, G. Duinkerken, R. de Jong, P.J. Van den Elsen, J.C. Hutton, S. Martin, B.O. Roep, and R.R.P. De Vries, 1997. Th1-like cytokine production profile and individual specific alterations in TCRBV-gene usage of T-cells from newly diagnosed type 1 diabetes patients after stimulation with β -cell autoantigens. *J Autoimmun* **10**, 589-598.
22. Bougneres, P.F., J.C. Carel, L. Castano, C. Boitard, J.P. Gardin, P. Landais et al., 1988. Factors associated with early remission of type I diabetes in children treated with cyclosporine. *N. Engl. J. Med.* **318**:663-670.
23. Harrison, L.C., P.G. Colman, B. Dean, R. Baxter and F.I.R. Martin, 1985. Increase in remission rate in newly diagnosed type 1 diabetic subjects treated with azathioprine. *Diabetes* **34**:1306-1308.
24. Lampeter, E.F., M. Homberg, K. Quabeck, U.W. Schaefer, P. Wernet, J. Bertrams et al., 1993. Transfer of insulin-dependent diabetes between HLA-identical siblings by bone marrow transplantation. *Lancet* **341**:1243-1244.
25. Sibley, R., D.E.R. Sutherland, F. Goetz and A.F. Michael, 1985. Recurrent diabetes mellitus in the pancreas iso- and allograft. A light and electron microscopic and immunohistochemical analysis of four cases. *Lab. Invest.* **53**:132-144.
26. Bingley, P.J., E. Bonifacio and E.A. Gale, 1993. Can we really predict IDDM?. [Review]. *Diabetes* **42**:213-220.
27. Baekkeskov, S., H.J. Aanstoot, S. Christgau, A. Reetz, M. Solimena, M. Cascalho et al., 1990. Identification of the 64K autoantigen in insulin-dependent diabetes as the GABA-synthesizing enzyme glutamic acid decarboxylase. *Nature* **347**:151-156.
28. Atkinson, M.A., D.L. Kaufman, L. Campbell, K.A. Gibbs, S.C. Shah, D.F. Bu et al., 1992. Response of peripheral-blood mononuclear cells to glutamate decarboxylase in insulin-dependent diabetes. *Lancet* **339**:458-459.
29. Palmer, J.P., C.M. Asplin, P. Clemons, K. Lyen, O. Tatpati, P.K. Raghu et al., 1983. Insulin antibodies in insulin-dependent diabetes before insulin treatment. *Science* **222**:1337-1339.
30. Schloot, N.C., B.O. Roep, D. Wegmann, L. Yu, H.P. Chase, T. Wang et al., 1997. Altered immune response to insulin in newly diagnosed versus insulin treated diabetic patients and healthy control subjects. *Diabetologia* **40**:564-572.

31. Pietropaolo, M., L. Castano, S. Babu, R. Buelow, Y.L. Kuo, S. Martin et al., 1993. Islet cell autoantigen 69 kD (ICA69). Molecular cloning and characterization of a novel diabetes-associated autoantigen. *J. Clin. Invest.* **92**:359-371.
32. Roep, B.O., G. Duinkerken, G.M. Schreuder, H. Kolb, R.R. de Vries and S. Martin, 1996. HLA-associated inverse correlation between T cell and antibody responsiveness to islet autoantigen in recent-onset insulin-dependent diabetes mellitus. *Eur J Immunol* **26**:1285-1289.
33. Arden, S.D., B.O. Roep, P.I. Neophytou, E.F. Usac, G. Duinkerken, R.R. de Vries et al., 1996. Imogen 38: a novel 38-kD islet mitochondrial autoantigen recognized by T cells from a newly diagnosed type 1 diabetic patient. *J Clin Invest* **97**:551-561.
34. Roep, B.O., S.D. Arden, R.R.P. De Vries and J.C. Hutton, 1990. T-cell clones from a type-1 diabetes patient respond to insulin secretory granule proteins. *Nature* **345**:632-634.
35. Bottazzo, G.F., B.M. Dean, J.M. McNally, E.H. MacKay, P.G.F. Swift and D.R. Gamble, 1985. In Situ Characterization of Autoimmune Phenomena and expression of HLA Class II Molecules in the Pancreas in Diabetic Insulinitis. *N. Engl. J. Med.* **313**:353-360.
36. Fujita, T.R., Y. Yui, Y. Kusumoto, Y. Serizawa, S. Makino and Y. Tochino, 1982. Lymphocytic insulinitis in a 'non-obese diabetic strain' of mice: an immunohistochemical and electron microscope investigation. *Biomed. Res.* **3**:429-429.
37. Atkinson, M.A., N.K. Maclaren and R. Luchetta, 1990. Insulinitis and diabetes in NOD mice reduced by prophylactic insulin therapy. *Diabetes* **39**:933-937.
38. Burkart, V. and H. Kolb, 1993. Protection of Islet Cells from Inflammatory Cell Death In vitro. *Clin. Exp. Immunol.* **93**:273-278.
39. Colucci, F., U. Dahl, L. Oreilly, A. Cooke, P. Chandler, E. Simpson et al., 1994. Non-diabetogenic insulinitis in NOD<->B10.GD allophenic mice in spite of permissive class I MHC antigens. *Scand. J. Immunol.* **40**:659-664.
40. Katz, J.D., B. Wang, K. Haskins, C. Benoist, D. Mathis, 1993. Following a Diabetogenic T-Cell from Genesis Through Pathogenesis. *Cell* **74**:1089-1100.
41. Foulis, A.K., M. McGill and M.A. Farquharson, 1991. Insulinitis in Type-1 (Insulin-Dependent) Diabetes-Mellitus in Man – Macrophages, Lymphocytes, and Interferon-gamma Containing Cells. *J. Pathol.* **165**:97-103.
42. Somoza, N., F. Vargas, C. Rouramir, M. Vivespi, M.T. Fernandezfigueras, A. Ariza et al., 1994. Pancreas in recent onset insulin-dependent diabetes mellitus – Changes in HLA, adhesion molecules and autoantigens, restricted T cell receptor V beta usage, and cytokine profile. *J. Immunol.* **153**:1360-1377.
43. Bendelac, A., C. Boitard, P. Bedossa, H. Bazin, J.F. Bach and C. Carnaud, 1988. Adoptive T cell transfer of autoimmune nonobese diabetic mouse diabetes does not require recruitment of host B lymphocytes. *J. Immunol.* **141**:2625-2628.
44. Wicker, L.S., B.J. Miller and Y. Mullen, 1986. Transfer of autoimmune diabetes mellitus with splenocytes from nonobese diabetic (NOD) mice. *Diabetes* **35**:855-860.
45. Haskins, K. and M. McDuffie, 1990. Acceleration of diabetes in young NOD mice with a CD4+ islet-specific T cell clone. *Science* **249**:1433-1436.
46. Chatenoud, L., E. Thervet, J. Primo and J.F. Bach, 1994. Anti-CD3 Antibody Induces Long-Term Remission of Overt Autoimmunity in Nonobese Diabetic Mice. *Proc. Natl. Acad. Sci. USA* **91**:123-127.
47. Kaufman, D.L., M.J. Clare-Salzler, J.D. Tian, T. Forsthuber, G.S.P. Ting, P. Robinson et al., 1993. Spontaneous loss of T-cell tolerance to glutamic acid decarboxylase in murine insulin-dependent diabetes. *Nature* **366**:69-72.
48. Wegmann, D.R., M. Norbury-Glaser and D. Daniel, 1994. Insulin-specific T cells are a predominant component of islet infiltrates in pre-diabetic NOD mice. *Eur J Immunol* **24**:1853-1857.
49. Schloot, N.C., S. Willems, G. Duinkerken, R.R.P. De Vries, B.O. Roep, 1998. Cloned T cells from a recent onset IDDM patient reactive with insulin B-chain. *J. Autoimmun.* **11**:169-175.
50. Healey, D., P. Ozegbe, S.D. Arden, P. Chandler, J.C. Hutton and A. Cooke, 1995. In vivo activity and in vivo specificity of CD4+ Th1 and Th2 cells derived from the spleens of diabetic NOD mice. *J. Clin. Invest.* **95**:2979-2985.
51. Roep, B.O., A.A. Kallan, W.L. Hazenbos, G.J. Bruining, E.M. Bailyes, S.D. Arden et al., 1991. T-cell reactivity to 38 kD insulin-secretory-granule protein in patients with recent-onset type 1 diabetes. *Lancet* **337**:1439-1441.
52. Boitard, C., M.C. Villa, C. Becourt, H.P. Gia, C. Huc, P. Sempe et al., 1992. Peripherin – An Islet Antigen That Is Cross-Reactive with Nonobese Diabetic Mouse Class-II Gene Products. *Proc. Natl. Acad. Sci. USA* **89**:172-176.
53. Elias, D., D. Markovits, T. Reshef, R. Van der Zee and I.R. Cohen, 1990. Induction and therapy of autoimmune diabetes in the non-obese diabetic (NOD/Lt) mouse by a 65-kDa heat shock protein. *Proc. Natl. Acad. Sci. USA* **87**:1576-1580.

54. Ellis, T.M., D.A. Schatz, E.W. Ottendorfer, M.S. Lan, C. Wasserfall, P.J. Salisburly et al., 1998. The relationship between humoral and cellular immunity to IA-2 in IDDM. *Diabetes* **47**:566-569.
55. Patel, S.D., A.P. Cope, M. Congia, T.T. Chen, E. Kim, L. Fugger et al., 1997. Identification of immunodominant T cell epitopes of human glutamic acid decarboxylase 65 by using HLA-DR (alpha *0101, beta* 0401)transgenic mice. *Proc Natl Acad Sci U S A* **94**:8082-8087.
56. Wen, L., S.Wong, L. Burkly, M. Altieri, C. Mamalaki, D. Kioussis, R.A. Flavell and R.S. Sherwin, 1998. Induction of insulinitis by glutamic acid decarboxylase peptide-specific HLA-DQ8-restricted T-cells from human DQ transgenic mice. *J. Clin. Invest.* **102**, 947-957.
57. Wicker, L.S., S.L. Chen, G.T. Nepom, J.F. Elliott, D.C. Freed, A. Bansal et al. Naturally processed T cell epitopes from human glutamic acid decarboxylase identified using mice transgenic for the type 1 diabetes-associated human MHC class II allele, DRB1*0401. *J Clin Invest* 1996. **98**:2597-2603.
58. Endl, J., H. Otto, G. Jung, B. Dreisbusch, F. Donie, P. Stahl et al., 1997. Identification of naturally processed T cell epitopes from glutamic acid decarboxylase presented in the context of HLA-DR alleles by T lymphocytes of recent onset IDDM patients. *J Clin Invest* **99**:2405-2415.
59. Geluk A., K.E. Van Meijgaarden, N.C. Schloot, J.W. Drijfhout, T.H.M. Ottenhoff and B.O. Roep, 1998. HLA-DR binding motifs of peptides from islet antigens in insulin-dependent diabetes mellitus. *Diabetes* **47**:1594-1601.
60. Hiemstra, H.S., P.A. Van Veelen, N.C. Schloot, K.E. Van Meijgaarden, S.J.M. Willemsen, J.A.M. Leunissen, W.E. Benckhuijsen, R. Amons, R.R.P. De Vries, B.O. Roep, T.H.M. Ottenhoff and J.W. Drijfhout, 1998. Definition of natural T-cell antigens with mimicry epitopes obtained from dedicated synthetic peptide libraries. *J. Immunol.* **161**, 4078-4082.
61. Bach, J.F., J. Dupre, G.S. Eisenbarth, L.C. Harrison, N.K. Maclaren, J. Nerup et al., 1990. Immunotherapy in pre-type 1 diabetes mellitus letter. *Diabetologia* **33**:741-741.
62. Muir, A., D.A. Schatz, P. Pozzilli and N.K. Maclaren, 1993. Intervention Therapies for Insulin-Dependent Diabetes. *Autoimmunity* **16**:301-310.
63. Pozzilli, P. and H. Kolb, 1989. Immunotherapy in type 1 diabetes: present status [news]. *Immunol. Today* **10**:321-322.
64. Elliott, R.B., C.C. Pilcher, A. Stewart, D. Fergusson and M.A. McGregor, 1993. The Use of Nicotinamide in the Prevention of Type 1 Diabetes. *Immunosuppressive. and Antiinflammatory. Drugs* **696**. -341
65. Schloot, N. and G.S. Eisenbarth, 1995. Isohormonal therapy of endocrine autoimmunity. *Immunology Today* **16**:289-294.
66. Maclaren, N.K., 1993. Diabetes intervention therapy. [Review]. *J. Diabet. Compl.* **7**:151-156.
67. Zamvil, Z.J., L. Davidson, G. Eisenbarth and H.L. Weiner, 1991. Suppression of diabetes in non-obese diabetic mice by oral administration of porcine insulin. *Proc Natl Acad Sci U S A* **88**:10252-10256.
68. Harrison, L.C., M. Domspey-Collier, D.R. Kramer, and K. Takahashi, 1996. Aerosol insulin induces regulatory CD8 gamma/delta T-cells that prevent murine insulin-dependent diabetes. *J. Exp. Med.* **184**, 2167-2174.
69. Tisch, R., X.D. Yang, S.M. Singer, R.S. Liblau, L. Fugger and H.O. McDevitt, 1993. Immune response to glutamic acid decarboxylase correlates with insulinitis in non-obese diabetic mice. *Nature* **366**:72-75.
70. Haskins, K. and D. Wegmann, 1996. Diabetogenic T-cell clones. *Diabetes* **45**:1299-1305.
71. Tian, J., M.A. Atkinson, M. Clare-Salzler, A. Herschenfeld, T. Forsthuber, P.V. Lehmann et al., 1996. Nasal administration of glutamate decarboxylase (GAD65) peptides induces Th2 responses and prevents murine insulin-dependent diabetes. *J. Exp. Med.* **183**:1561-1567.
72. Tian, J., M. Clare-Salzler, A. Herschenfeld, B. Middleton, D. Newman, R. Mueller et al., 1996. Modulating autoimmune response to GAD inhibits disease progression and prolongs islet graft survival in diabetes-prone mice. *Nature Medicine* **12**:1348-1353.
73. Bowman, M.A., E.H. Leiter and M.A. Atkinson, 1994. Prevention of Diabetes in the Nod Mouse – Implications for Therapeutic Intervention in Human Disease. *Immunol. Today* **15**:115-120.
74. Daniel, D. and D. Wegmann, 1996. Protection of nonobese diabetic mice from diabetes by intranasal or subcutaneous administration of insulin peptide B-(9-23). *Proc. Natl. Acad. Sci. USA* **93**:956-960.
75. Zhang, J., Davidson L, Eisenbarth GS, Weiner H. Suppression of diabetes in non-obese diabetic mice by oral administration of porcine insulin. *Proc. Natl. Acad. Sci. USA* 1991. **88**:10252-10256.
76. Gerling, I.C., M.A. Atkinson, A.B. Peck, J.G. Cornelius and E.H. Leiter, 1994. Intrathymic (IT) Injection of a Glutamic Acid Decarboxylase (GAD) derived Peptide Accelerates Diabetes Onset in NOD/Lt Females. *Diabetes* **43**:93A.

77. Allison, J., L.C. Harrison, I.L. Campbell and J.F. Miller, 1990. Major histocompatibility complex molecules and the beta cell: inferences from transgenic models. *Curr. Top. Microbiol. Immunol.* **156**:121-135.
78. Ohashi, P.S., S. Oehen, K. Buerki, H. Pirchner, C.T. Ohashi, B. Odermatt et al., 1991. Ablation of 'Tolerance' and Induction Of Diabetes by Virus Infection in Viral Antigen Transgenic Mice. *Cell* **65**:305-317.
79. Oreilly, L.A., D. Healey, E. Simpson, P. Chandler, T. Lund, M.A. Ritter et al., 1994. Studies on the thymus of non-obese diabetic (NOD) mice: Effect of transgene expression. *Immunology* **82**:275-286.
80. Sarvetnick, N., 1990. Transgenic models of diabetes. *Curr. Opin. Immunol.* **2**:604-606.
81. Allison, J., L. Oxbrow and J.F.A.P. Miller, 1994. Consequences of in Situ Production of IL-2 for Islet Cell Death. *Int. Immunol.* **6**:541-549.
82. Stewart, T.A., B. Hultgren, X. Huang, S. Pittsmeek, J. Hully and N.J. MacLachlan, 1993. Induction of Type-I Diabetes by Interferon-alpha in Transgenic Mice. *Science* **260**:1942-1946.
83. Campbell, I.L., M.V. Hobbs, J. Dockter, M.B. Oldstone and J. Allison, 1994. Islet inflammation and hyperplasia induced by the pancreatic islet-specific overexpression of interleukin-6 in transgenic mice. *Am. J. Pathol.* **145**:157-166.
84. Guerder, S., D.E. Picarella, P.S. Linsley and R.A. Flavell, 1994. Costimulator B7-1 confers antigen-presenting-cell function to parenchymal tissue and in conjunction with tumor necrosis factor alpha leads to autoimmunity in transgenic mice. *Proc. Natl. Acad. Sci. USA* **91**:5138-5142.
85. Wogensen, L., M.S. Lee and N. Sarvetnick, 1994. Production of Interleukin-10 by Islet Cells Accelerates Immune-Mediated Destruction of beta-Cells in Nonobese Diabetic Mice. *J. Exp. Med.* **179**:1379-1384.
86. Dalton, D.K., S. Pittsmeek, S. Keshav, I.S. Figari, A. Bradley and T.A. Stewart, 1993. Multiple Defects of Immune Cell Function in Mice with Disrupted Interferon-gamma Genes. *Science* **259**:1739-1742.
87. Gores, P.F., J.L. Platt, F. Rabe, P. Stock and D.E. Sutherland, 1987. Permanent acceptance of islet allografts in mice is not associated with immunologic tolerance. *Transplant. Proc.* **19**:499-500.
88. Bradley, B.J., K. Haskins, F.G. Larosa and K.J. Lafferty, 1992. CD8 T-Cells Are Not Required for Islet Destruction Induced by a CD4+ Islet-Specific T-Cell Clone. *Diabetes* **41**:1603-1608.
89. Hao, L., Y. Wang, R.G. Gill and K.J. Lafferty, 1987. Role of the L3T4+ T cell in allograft rejection. *J. Immunol.* **139**:4022-4026.
90. Posselt, A.M., C.F. Barker, J.E. Tomaszewski, J.F. Markmann, M.A. Choti and A. Naji, 1990. Induction of donor-specific unresponsiveness by intrathymic islet transplantation. *Science* **249**:1293-1295.

Dr. B.O. Roep
 Dept. Immunohaematology & Blood Bank
 Leiden University Medical Centre
 P.O.Box 9600
 2300 RC Leiden, the Netherlands
 tel: +31-71-263869
 fax: +31-71-5216751
 e-mail: broep@pobox.LeidenUniv.NL

